



Petrifilm™

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3M™ Petrifilm™
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3M™ Petrifilm™ Yeast and Mold Count Plate

Performance Summary

3M™ Petrifilm™ Yeast and Mold Count Plates are sample-ready media plates used for the enumeration of yeast and mold in the food and beverage industries. Each plate contains a water-soluble gelling agent, nutrients and indicators in a dry, shelf-stable format.

This technical bulletin summarizes data 3M Food Safety collected during performance testing of 3M Petrifilm Yeast and Mold Count Plates for microbial testing of bottled water.

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3M Petrifilm Yeast and Mold Count Plate Performance Data

Comparative Method Study

A method comparison study was conducted at an external reference laboratory to compare results of the 3M Petrifilm Yeast and Mold Count Plate method to a reference method, Chlortetracycline Rose Bengal¹ (CRB) agar and to another agar, acidified Potato Dextrose Agar (aPDA), for the recovery of yeast and mold in bottled water.

Matrices: Fifteen brands of water were tested, two lots per brand to equal a total of 30 samples.

Table 1: Brands of bottled water tested

| Water Type | Brand, Country of Manufacture | Water Type | Brand, Country of Manufacture |
|-----------------|---|-----------------------------|--|
| Purified | Nestle Pure Life, United States Aquafina, United States Dasani, United States | Natural Spring | Evian, France Jana, Croatia Fiji, Fiji Ty Nant, Wales Voss Flat, Norway |
| Regional Spring | Trauth Dairy, United States Ice Mountain, United States Kroger, United States | Natural Spring (Carbonated) | Voss Sparkling, Norway Gerolsteiner, Germany Apollinaris, Germany Perrier, France |

Organisms

Overnight broth cultures were washed and inoculated into bottled water samples at a low/medium inoculum level (targeting 25 cfu/sample) and a medium/high inoculum level (targeting 75 cfu/sample). A non-inoculated control was also prepared for each water sample.

The following three organisms were randomly assigned to the 30 different water types: *Penicillium sp.* ATCC #18307, *Paecilomyces sp.* ATCC #1114, and *Candida albicans* ATCC #10231. Inoculated water was left in the dark at room temperature overnight to equilibrate before filtering.

¹ Standard Methods for Examination of Water and Wastewater (SMEWW) method 9610D 2a - Membrane filter technique for detection of fungi, 20th Edition, 1998.

3M Petrifilm Yeast and Mold Count Plate Performance Data

Comparative Method Study (continued)

Method Comparison

The following yeast and mold methods were compared:

- ▶ 3M Petrifilm Yeast and Mold Count Plates vs. Chlortetracycline Rose Bengal [CRB] agar (100 mL filtered; SMEWW). Plates were incubated at $20 \pm 1^\circ\text{C}$ and counted at 3 & 5 days.
- ▶ 3M Petrifilm Yeast and Mold Count Plates vs. acidified Potato Dextrose [aPDA] agar (100 mL filtered; customer method). Plates were incubated at $25 \pm 1^\circ\text{C}$ and counted at 3 & 5 days.

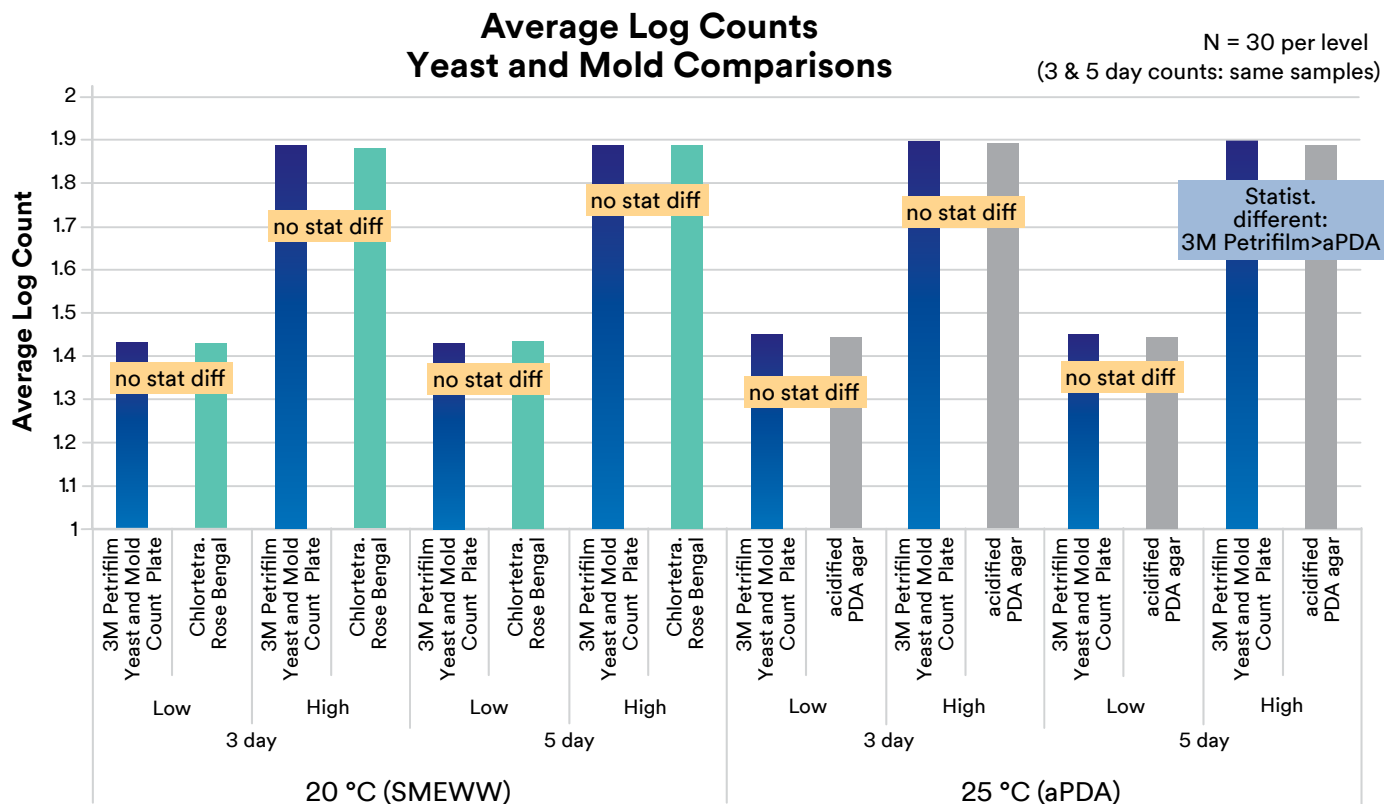
A mixed cellulose ester filter was used for the method comparison. Samples were plated in duplicate for each level of inoculum. After incubation at the conditions noted above, all colonies on all plates were counted.

Statistical Analysis

Counts were converted to \log_{10} counts. A paired t-test per inoculation level was used to compare differences in counts between the 3M Petrifilm Yeast and Mold Count Plate method and the comparative methods. A p-value of <0.05 was taken to indicate a significant difference.

Results

Graph 1: Yeast and Mold Comparisons: Average Log Counts



SMEWW method comparison

- ▶ The mean log yeast and mold counts were not significantly different between the 3M Petrifilm Yeast and Mold Count Plate method and the CRB agar method at the low inoculation level at day 3 (p-value=0.745) or the high inoculation level at day 3 (p-value=0.063).
- ▶ The mean log yeast and mold counts were not significantly different between the 3M Petrifilm Yeast and Mold Count Plate method and the CRB agar method at the low inoculation level at day 5 (p-value=0.121) or the high inoculation level at day 5 (p-value=0.052).

Customer method comparison

- ▶ The mean log yeast and mold counts were not significantly different between the 3M Petrifilm Yeast and Mold Count Plate method and the aPDA method at the low inoculation level at day 3 (p-value=0.844) or the high inoculation level at day 3 (p-value=0.723).
- ▶ The mean log yeast and mold counts were not significantly different between the 3M Petrifilm Yeast and Mold Count Plate method and the aPDA method at the low inoculation level at day 5 (p-value=0.506).
- ▶ At the high inoculation level at day 5, the mean log yeast and mold counts per filter were significantly different between the 3M Petrifilm Yeast and Mold Count Plate method and the aPDA method (p=0.036), with the log counts per filter for the 3M Petrifilm Yeast and Mold Count Plate method, on average, greater in value than those from the aPDA method.

3M Petrifilm Yeast and Mold Count Plate Performance Data

Inclusivity/Exclusivity Study

Inclusivity and exclusivity studies were performed in an external and internal study that combined consists of 19 fungal and 19 non-fungal pure culture strains. The strains were obtained from the American Type Culture Collection (USA) or from the 3M culture collection. Fungal organisms were diluted and inoculated into bottled water targeting 25-50 cfu/filter; non-fungal strains were diluted and inoculated targeting approximately 1×10^5 cfu/filter.

Inoculated samples were filtered through a mixed cellulose ester filter and plated onto 3M Petrifilm Yeast and Mold Count Plates, Chlorotetracycline Rose-Bengal agar (CRB) or acidified potato dextrose agar (aPDA).

1. One set of 3M Petrifilm Yeast and Mold Count Plates and the CRB plates were incubated at 20 ± 1 °C for 5 days, reading at both 3 and 5 days.
2. A second set of 3M Petrifilm Yeast and Mold Count Plates and the aPDA plates were incubated at 25 ± 1 °C for 5 days, reading at both 3 and 5 days.
3. Non-fungal strains were also filtered and plated onto plate count agar and incubated at 35 ± 1 °C for 48 ± 2 h as a positive control.

Inclusivity Strains

| | |
|-------------------------------------|-------------|
| <i>Alternaria alternaria</i> | ATCC 6663 |
| <i>Aspergillus brasiliensis</i> | ATCC 16404 |
| <i>Aspergillus niger</i> | M6 (3M) |
| <i>Candida albicans</i> | ATCC 10231 |
| <i>Candida albicans</i> | ATCC 66027 |
| <i>Candida glabrata</i> | ATCC 26512 |
| <i>Candida guilliermondii</i> | ATCC 6260 |
| <i>Chaetomium globosum</i> | ATCC 6205 |
| <i>Cladosporium cladosporioides</i> | ATCC16022 |
| <i>Hansenula (Pichia) anomala</i> | ATCC 2349 |
| <i>Paecilomyces variotii</i> | ATCC 1114 |
| <i>Penicillium corylophilum</i> | ATCC 18307 |
| <i>Pityrosporum ovale</i> | ATCC 12078 |
| <i>Rhizopus stolonifer</i> (+) | ATCC 6227b |
| <i>Rhodotorula mucilaginosa</i> | ATCC 9449 |
| <i>Saccharomyces cerevisiae</i> | ATCC 18824 |
| <i>Saccaromyces cerevisiae</i> | ATCC 2601 |
| <i>Scopulariopsis acremonium</i> | ATCC 58636 |
| <i>Trichosporon mucoides</i> | ATCC 204094 |

Exclusivity Strains

| | |
|--|------------|
| <i>Acinetobacter baumannii</i> | ATCC 19606 |
| <i>Aeromonas hydrophila</i> | ATCC 7965 |
| <i>Bacillus atrophaeus</i> | ATCC 51189 |
| <i>Bacillus cereus</i> | ATCC 11774 |
| <i>Bacillus pumilis</i> | ATCC 14884 |
| <i>Enterobacter aerogenes</i> | ATCC 13048 |
| <i>Enterococcus faecalis</i> | ATCC 29212 |
| <i>Escherichia coli</i> | ATCC 25922 |
| <i>Enterococcus faecalis</i> | ATCC 29212 |
| <i>Escherichia coli</i> | ATCC 25922 |
| <i>Lactobacillus delbrueckii subsp. lactis</i> | ATCC 4797 |
| <i>Micrococcus luteus</i> | ATCC 10240 |
| <i>Pseudomonas aeruginosa</i> | ATCC 15442 |
| <i>Pseudomonas fluorescens</i> | ATCC 13525 |
| <i>Salmonella enteritidis typhimurium</i> | ATCC 13311 |
| <i>Salmonella enterica</i> | ATCC 51812 |
| <i>Sphingomonas paucimobilis</i> | ATCC 29837 |
| <i>Staphylococcus aureus</i> | ATCC 25923 |
| <i>Yersinia enterocolitica</i> | ATCC 9610 |

Results Comparison to SMEWW Method, 20 °C

Inclusivity

| | 3M Petrifilm Yeast and Mold Count Plates | CRB Agar |
|--------|---|---|
| | 15/19 strains (79%) had growth | 15/19 strains (79%) had growth |
| 3 days | Strains with no visible growth: <i>Alternaria alternaria</i> ATCC 6663 <i>Chaetomium globosum</i> ATCC 6205 <i>Cladosporium cladosporioides</i> ATCC 16022 <i>Scopulariopsis acremonium</i> ATCC 58636 | Strains with no visible growth: <i>Alternaria alternaria</i> ATCC 6663 <i>Chaetomium globosum</i> ATCC 6205 <i>Cladosporium cladosporioides</i> ATCC 16022 <i>Scopulariopsis acremonium</i> ATCC 58636 |
| 5 days | 19/19 strains (100%) had growth | 19/19 strains (100%) had growth |

Exclusivity

| | 3M Petrifilm Yeast and Mold Count Plates | CRB Agar |
|--------|---|--|
| | 4/19 strains (21%) had growth | 1/19 strains (5%) had growth |
| 5 days | Strains with visible growth: <i>Acinetobacter baumannii</i> ATCC 19606 (1 colony) <i>Aeromonas hydrophila</i> ATCC 7965 <i>Enterobacter aerogenes</i> ATCC 13048 (1 colony) <i>Enterococcus faecalis</i> ATCC 29212 (1 colony) | Strains with visible growth: <i>Aeromonas hydrophila</i> ATCC 7965 |

Results Comparison to Customer Method, 25 °C

Inclusivity

| | 3M Petrifilm Yeast and Mold Count Plates | aPDA Agar |
|--------|---|---|
| | 5/19 strains (79%) had growth | 15/19 strains (79%) had growth |
| 3 days | Strains with no visible growth: <i>Alternaria alternaria</i> ATCC 6663 <i>Chaetomium globosum</i> ATCC 6205 <i>Cladosporium cladosporioides</i> ATCC 16022 <i>Scopulariopsis acremonium</i> ATCC 58636 | Strains with no visible growth: <i>Alternaria alternaria</i> ATCC 6663 <i>Chaetomium globosum</i> ATCC 6205 <i>Cladosporium cladosporioides</i> ATCC 16022 <i>Scopulariopsis acremonium</i> ATCC 58636 |
| 5 days | 20/20 strains (100%) had growth | 18/19 strains (95%) had growth |
| | | Strains with no visible growth: <i>Scopulariopsis acremonium</i> ATCC 58636 |

Exclusivity

| | 3M Petrifilm Yeast and Mold Count Plates | aPDA Agar |
|--------|---|--|
| | 4/19 strains (21%) had growth | 1/19 strains (5%) had growth |
| 5 days | Strains with visible growth: <i>Aeromonas hydrophila</i> ATCC 7965 <i>Bacillus atrophaeus</i> ATCC 51189 (1 colony) <i>Bacillus cereus</i> ATCC 11774 (1 colony) <i>Salmonella enteritidis</i> ATCC 13311 (1 colony) | Strains with visible growth: <i>Aeromonas hydrophila</i> ATCC 7965 |



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